



**Visualisation of bio-interaction between noble metals' nanoparticles and bacteria  
*Salmonella Enterididis* (G-) and *Listeria monocytogenes* (G+)**

Sawosz, Ewa; Niemiec, Tomasz; Szeliga, J.; Chwalibog, André; Sawosz, Filip; Sokolowska, Alexander; Krajewska-Kaminska, E.

*Published in:*  
Abstracts Book

*Publication date:*  
2009

*Document version*  
Publisher's PDF, also known as Version of record

*Citation for published version (APA):*  
Sawosz, E., Niemiec, T., Szeliga, J., Chwalibog, A., Sawosz, F., Sokolowska, A., & Krajewska-Kaminska, E. (2009). Visualisation of bio-interaction between noble metals' nanoparticles and bacteria *Salmonella Enterididis* (G-) and *Listeria monocytogenes* (G+). In *Abstracts Book*

4th International Conference on Surfaces,  
Coatings and Nanostructured Materials  
(NANOSMAT 2009)

# ABSTRACTS BOOK

Editors:

Dr N. Ali (UK) & Dr R. Polini (Italy)

**NANOSMAT 2009 (19th-22th October 2009)**

4th International Conference on Surfaces, Coatings and Nanostructured Materials

Crown Plaza Hotel,

Via Aurelia Antica 415

00165 Rome

**N571: Visualisation of bio-interaction between noble metals' nanoparticles and bacteria *Salmonella Enteritidis* (G-) and *Listeria monocytogenes* (G+)**

E. Sawosz<sup>1</sup>, T. Niemiec<sup>1</sup>, J. Szeliga, A., Chwalibog<sup>2</sup>, F. Sawosz<sup>1,2</sup>, A. Sokołowska<sup>3</sup>, E. Krajewska-Kamińska

<sup>1</sup>University of Life Science, Ciszewskiego 8, 02-786 Warsaw, Poland

<sup>2</sup>University of Copenhagen, Groennegaardsvej 3, 1870 Frederiksberg, Denmark

230

<sup>3</sup>Technical University of Warsaw, Plac Politechniki 1, 02-780 Warsaw, Poland

The objective of experiments was to assess morphology of bio-relation of noble metals' nanoparticles (Cu, Ag, Au, and Pt) and bacteria, visualized with transmission electron microscope. Hydrocolloids of Cu, Ag, Au and Pt, obtained from Nano-Tech, Poland were produced by electric non-explosive method (Polish patent 380649). Pure bacteria: *Salmonella Enteritidis* (G-) and *Listeria monocytogenes* (G+) were dissolved in redistilled water. Immediately after bacteria were aliquoted, they were mixed into test-tubes with hydrocolloids of Cu, Ag, Au, and Pt nanoparticles (each at a concentration of 50 ppm), in proportion 1:1. Samples of Cu, Ag, Au and Pt, solutions of nanoparticles with bacteria and pure bacteria as controls were prepared by placing droplets on copper, formvaried grid (Agar Scientific Ltd., Stansted, UK). Immediately after drying of the droplets in dry air, grids were inserted into transmission electron microscope (TEM) (JEOL model JEM-2000EX). Morphology of bacteria and localisation of nanoparticles were inspected by TEM. *Salmonella* (G-) and *Listeria* (G+), as well as type of metals affected obtained pictures. Affinity of Ag to cell's wall (seen as a chain of nanoparticles located around each cell) of *Listeria* but not *Salmonella* was demonstrated. Also some nanoparticles were seen inside *Salmonella* but not *Listeria*. Gold nanoparticles were aggregated and surrounded by cells of *Listeria* but not by *Salmonella*. Effect of Pt nanoparticle on *Listeria* and *Salmonella* morphology was similar, showing release of mesosomes from bacteria cell's. Cu nanoparticles destroyed cell's wall, and also were attached to biofilm of bacteria. The present results indicate that both distribution and behaviour of nanoparticles depend on metals and kind of bacteria exposed to the nanoparticles.